Package ‘fishpond’

October 16, 2019

Title Fishpond: differential transcript and gene expression with inferential replicates

Version 1.0.2

Author Anqi Zhu, Avi Srivastava, Joseph Ibrahim, Rob Patro, Michael Love

Maintainer Michael Love <michaelisaiahlove@gmail.com>

Description Fishpond contains methods for differential transcript and gene expression analysis of RNA-seq data using inferential replicates for uncertainty of abundance quantification, as generated by Gibbs sampling or bootstrap sampling.

Imports graphics, stats, utils, methods, abind, gtools, qvalue, S4Vectors, SummarizedExperiment, matrixStats, svMisc

Suggests testthat, knitr, rmarkdown, macrophage, tximeta, org.Hs.eg.db, samr, DESeq2, apeglm

License GPL-2

Encoding UTF-8

URL https://github.com/mikelove/fishpond

biocViews Sequencing, RNASeq, GeneExpression, Transcription, Normalization, Regression, MultipleComparison, BatchEffect, Visualization, DifferentialExpression, DifferentialSplicing, AlternativeSplicing, SingleCell

VignetteBuilder knitr

LazyData true

RoxygenNote 6.1.1

git_url https://git.bioconductor.org/packages/fishpond

git_branch RELEASE_3_9

git_last_commit c1c84a3

git_last_commit_date 2019-08-11

Date/Publication 2019-10-15

R topics documented:

deswish 2
labelKeep 3

makeSimSwishData 3
plotInfReps 4
plotMASwish 5
scaleInfReps 6
swish 7

Index **9**

`deswish` *deswish: DESeq2-apeglm With Inferential Samples Helps*

Description

The DESeq2-apeglm With Inferential Samples implementation supposes a hierarchical distribution of log2 fold changes. The final posterior standard deviation is calculated by adding the posterior variance from modeling biological replicates computed by `apeglm`, and the observed variance on the posterior mode over inferential replicates. This function requires the DESeq2 and apeglm packages to be installed and will print an error if they are not found.

Usage

```
deswish(y, x, coef)
```

Arguments

`y`	a SummarizedExperiment containing the inferential replicate matrices, as output by `tximeta`, and then with `labelKeep` applied. One does not need to run `scaleInfReps` as scaling is done internally via DESeq2.
`x`	the design matrix
`coef`	the coefficient to test (see `lfcShrink`)

Value

a SummarizedExperiment with metadata columns added: the log2 fold change and posterior SD using inferential replicates, and the original log2 fold change (apeglm) and its posterior SD

References

The `DESeq` and `lfcShrink` function in the `DESeq2` package:

Zhu, Ibrahim, Love "Heavy-tailed prior distributions for sequence count data: removing the noise and preserving large differences" Bioinformatics (2018).

Love, Huber, Anders "Moderated estimation of fold change and dispersion for RNA-seq data with DESeq2" Genome Biology (2014).

Examples

```
y <- makeSimSwishData()
y <- labelKeep(y)
y <- deswish(y, ~condition, "condition_2_vs_1")
```

labelKeep — *Label rows to keep based on minimal count*

Description

Adds a column `keep` to `mcols(y)` that specifies which rows of the SummarizedExperiment will be included in statistical testing. Rows are not removed, just marked with the logical `keep`.

Usage

```
labelKeep(y, minCount = 10, minN = 3, x)
```

Arguments

`y`	a SummarizedExperiment
`minCount`	the minimum count
`minN`	the minimum sample size at `minCount`
`x`	the name of the condition variable, will use the smaller of the two groups to set `minN`. Similar to edgeR's `filterByExpr`, as the smaller group grows past 10, `minN` grows only by 0.7 increments of sample size

Value

a SummarizedExperiment with a new column `keep` in `mcols(y)`

Examples

```
y <- makeSimSwishData()
y <- scaleInfReps(y)
y <- labelKeep(y)
```

makeSimSwishData — *Make simulated data for swish for examples/testing*

Description

Makes a small swish dataset for examples and testing. The first six genes have some differential expression evidence in the counts, with varying degree of inferential variance across inferential replicates (1-2: minor, 3-4: some, 5-6: substantial). The 7th and 8th genes have all zeros to demonstrate `labelKeep`.

Usage

```
makeSimSwishData(m = 1000, n = 10, numReps = 20, null = FALSE)
```

Arguments

m	number of genes
n	number of samples
numReps	how many inferential replicates
null	logical, whether to make an all null dataset

Value

a SummarizedExperiment

Examples

```
library(SummarizedExperiment)
y <- makeSimSwishData()
assayNames(y)
```

---

plotInfReps *Plot inferential replicates for a gene or transcript*

Description

Plot inferential replicates for a gene or transcript

Usage

```
plotInfReps(y, idx, x, cov = NULL, cols.drk = c("dodgerblue",
  "goldenrod4"), cols.lgt = c("lightblue1", "goldenrod1"), xaxis)
```

Arguments

y	a SummarizedExperiment (see swish)
idx	the name or row number of the gene or transcript
x	the name of the condition variable
cov	the name of the covariate for adjustment
cols.drk	dark colors for the lines of the boxes
cols.lgt	light colors for the inside of the boxes
xaxis	logical, whether to label the sample numbers. default is TRUE if there are less than 30 samples

Value

nothing, a plot is displayed

Examples

```
y <- makeSimSwishData()
plotInfReps(y, 3, "condition")

y <- makeSimSwishData(n=40)
y$batch <- factor(rep(c(1,2,3,1,2,3),c(5,10,5,5,10,5)))
plotInfReps(y, 3, "condition", "batch")
```

---

plotMASwish *MA plot*

Description

MA plot

Usage

```
plotMASwish(y, alpha = 0.05, sigcolor = "blue", ...)
```

Arguments

y	a SummarizedExperiment (see swish)
alpha	the FDR threshold for coloring points
sigcolor	the color for the significant points
...	passed to plot

Value

nothing, a plot is displayed

Examples

```
y <- makeSimSwishData()
y <- scaleInfReps(y)
y <- labelKeep(y)
y <- swish(y, x="condition")
plotMASwish(y)
```

`scaleInfReps` *Scale inferential replicate counts*

Description

A helper function to scale the inferential replicates to the mean sequencing depth. The scaling takes into account a robust estimator of size factor (median ratio method is used). First, counts are corrected per row using the effective lengths (for gene counts, the average transcript lengths), then scaled per column to the geometric mean sequence depth, and finally are adjusted per-column up or down by the median ratio size factor to minimize systematic differences across samples.

Usage

```
scaleInfReps(y, lengthCorrect = TRUE, meanDepth = NULL, sfFun = NULL,
  minCount = 10, minN = 3, quiet = FALSE)
```

Arguments

`y`	a SummarizedExperiment with: `infReps` a list of inferential replicate count matrices, `counts` the estimated counts matrix, and `length` the effective lengths matrix
`lengthCorrect`	whether to use effective length correction (default is TRUE)
`meanDepth`	(optional) user can specify a different mean sequencing depth. By default the geometric mean sequencing depth is computed
`sfFun`	(optional) size factors function. An alternative to the median ratio can be provided here to adjust the scaledTPM so as to remove remaining library size differences
`minCount`	for internal filtering, the minimum count
`minN`	for internal filtering, the minimum sample size at `minCount`
`quiet`	display no messages

Value

a SummarizedExperiment wiht the inferential replicates as scaledTPM with library size already corrected (no need for further normalization)

Examples

```
y <- makeSimSwishData()
y <- scaleInfReps(y)
```

swish *swish: SAMseq With Inferential Samples Helps*

Description

swish: SAMseq With Inferential Samples Helps

Usage

```
swish(y, x, cov = NULL, pair = NULL, interaction = FALSE,
  nperms = 30, estPi0 = FALSE, qvaluePkg = "qvalue", pc = 5,
  nRandomPairs = 30, quiet = FALSE)
```

Arguments

y	a SummarizedExperiment containing the inferential replicate matrices of median-ratio-scaled TPM as assays 'infRep1', 'infRep2', etc.
x	the name of the condition variable. A factor with two levels for a two group analysis (possible to adjust for covariate or matched samples, see next two arguments)
cov	the name of the covariate for adjustment. If provided a stratified Wilcoxon in performed. Cannot be used with `pair`
pair	the name of the pair variable, which should be the number of the pair. Can be an integer or factor. If specified, a signed rank test is used to build the statistic. All samples across `x` must be pairs if this is specified. Cannot be used with `cov`.
interaction	logical, whether to perform a test of an interaction between `x` and `cov`. These are different than the other tests produced by the software, in that they focus on a difference in the log2 fold change across levels of `x` when comparing the two levels in `cov`. If `pair` is specified, this will perform a Wilcoxon rank sum test on the two groups of matched sample LFCs. If `pair` is not included, multiple random pairs of samples within the two groups are chosen, and again a Wilcoxon rank sum test compared the LFCs across groups.
nperms	the number of permutations
estPi0	logical, whether to estimate pi0
qvaluePkg	character, which package to use for q-value estimation, `samr` or `qvalue`
pc	pseudocount for finite estimation of `log2FC`, not used in calculation of test statistics, `locfdr` or `qvalue`
nRandomPairs	the number of random pseudo-pairs (only used with `interaction=TRUE` and un-matched samples) to use to calculate the test statistic
quiet	display no messages

Value

a SummarizedExperiment with metadata columns added: the statistic (either a centered Wilcoxon Mann-Whitney or a signed rank statistic, aggregated over inferential replicates), a log2 fold change (the median over inferential replicates, and averaged over pairs or groups (if groups, weighted by sample size), the local FDR and q-value, as estimated by the `samr` package.

References

swish is described in the following reference:

Anqi Zhu, Avi Srivastava, Joseph G Ibrahim, Rob Patro, Michael I Love "Nonparametric expression analysis using inferential replicate counts" Nucleic Acids Research (2019).

The swish method builds upon the SAMseq method, and extends it by incorporating inferential uncertainty. swish internally calls functions from the samr package, for example, the calculation of local FDR and q-value.

The citation for SAMseq is:

Jun Li and Robert Tibshirani "Finding consistent patterns: A nonparametric approach for identifying differential expression in RNA-Seq data" Stat Methods Med Res (2013).

Examples

```
library(SummarizedExperiment)
set.seed(1)
y <- makeSimSwishData()
y <- scaleInfReps(y)
y <- labelKeep(y)
y <- swish(y, x="condition")

# histogram of the swish statistics
hist(mcols(y)$stat, breaks=40, col="grey")
cols = rep(c("blue","purple","red"),each=2)
for (i in 1:6) {
  arrows(mcols(y)$stat[i], 20,
         mcols(y)$stat[i], 10,
         col=cols[i], length=.1, lwd=2)
}

# plot inferential replicates
plotInfReps(y, 1, "condition")
plotInfReps(y, 3, "condition")
plotInfReps(y, 5, "condition")
```

Index

deswish, 2

labelKeep, 3

makeSimSwishData, 3

plotInfReps, 4
plotMASwish, 5

scaleInfReps, 6
swish, 7